

*aggregata, globosa, ad 215µm diam.; ascospores oblongae vel ellipsoideae, 4-septatae, constrictae, raro cellula centralis magniora, 41-45 x 18-21µm.*

Colonies epiphyllous, dense, up to 2mm in diameter. Hyphae straight to substraight, branching opposite to irregular at acute angles, loosely to closely reticulate, cells 16-20 x 6-8µm. Appressoria alternate, about 2% opposite, antrorse to subantrorse, 16-21µm long; stalk cells cylindrical to cuneate, 3-7µm long; head cells ovate, clavate, globose, entire, 12-14 x 11-13µm. Phialides mixed with appressoria, alternate to opposite, ampulliform, 16-20 x 8-10µm. Mycelial setae few to numerous, simple, straight, about 2% curved to uncinuate, acute, dentate to cristate at the apex, up to 600µm long. Perithecia scattered to loosely grouped, globose, up to 215µm in diameter; ascospores oblong to ellipsoidal, 4-septate, constricted at the septa, rarely central cell larger, 41- 45 x 18-21µm.

### Remarks

Having acute to dentate mycelial setae, *M. smilacacearum* can be compared with *M. smilacis* Stev. known on *Smilax* spp. from Porto Rico, Honduras and Taiwan (Hansford, 1961) but differs from it in having only epiphyllous colonies, longer, straight but few uncinuate mycelial setae and smaller ascospores.

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## CASTRATION IN A COMMON MONGOOSE *HERPESTES EDWARDSI*

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*plus web supplement of 1 page*

Castration in non-domestic species is carried out to increase the docility of animals having intimate contact with humans (Fowler, 1985). The present paper describes open method of castration in a Common Mongoose (*Herpestes edwardsi*).

A male common mongoose, aged about six months and weighing 700g was presented to the surgery clinic of Orissa Veterinary College for castration. It was anaesthetized with a mixture of atropine sulphate 0.02mg and ketamine hydrochloride 7mg injected intramuscularly using an insulin syringe (Image 1<sup>w</sup>). The mongoose was recumbent within two minutes. A cover of hypodermic needle cut at its closed end was applied to both canine teeth to serve as mouth gag and the tongue was pulled out (Image 2<sup>w</sup>). After routine preparation of the site, castration was performed through scrotal incision as usual. Ligation of spermatic duct and vessels and transfixation and ligation of tunica vaginalis communis was done with 1-0 chromic cat gut. The skin was sutured with nylon suture after pouring povidone-iodine lotion (Image 3<sup>w</sup>). Cephalixin 125mg dispersible tablets (Ceff 15mg, DT. Lupin Lab, Mumbai) were administered orally once daily in fish soup for five days. Skin stitches were removed on the twelfth post-operative day and the animal recovered uneventfully.

Fowler (1985) opined that veterinarians must inform the client that castration may not necessarily decrease unpredictable behaviour or increase docility. In the present case castration of the mongoose was done due to the request of the owner inspite of the above suggestions being explained to him. The mongoose was well anaesthetized with the mixture of atropine sulphate and ketamine hydrochloride as suggested by Retting and Divers (1985) for restraint and handling of animals of family Viverridae.

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<sup>w</sup> See Images in the web supplement at [www.zoosprint.org](http://www.zoosprint.org)



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