

## TAXONOMIC STUDIES ON THE VEGETABLE PEST OF GENUS *PLUTELLA* SCHRÄNK (PLUTELLIDAE: LEPIDOPTERA) FROM NORTHWESTERN INDIA

P.C. Pathania<sup>1</sup> and H.S. Rose<sup>2</sup>

<sup>1</sup>Division of Entomology, Indian Agricultural Research Institute, New Delhi 110012, India

<sup>2</sup>Department of Zoology, Punjabi University, Patiala 147002, Punjab, India

Email: <sup>1</sup> pathaniapc@yahoo.co.in; <sup>2</sup> profhsrose@yahoo.com

web supplement

### ABSTRACT

*Taxonomic studies of Plutella xylostella Linnaeus from northwestern India was made based on the structural details of wing venation and external genitalia.*

### KEYWORDS

Genitalia, India, Lepidoptera, Plutellidae, Plutella

### ABBREVIATIONS

IA+2A - Vein representing fused first and second anal vein; 3A - Third anal vein; CuA<sub>1</sub> - First anterior cubital vein; CuA<sub>2</sub> - Second anterior cubital vein; CuP - Posterior cubital vein; FRI - Forest Research Institute; M<sub>1</sub> - First medial vein; M<sub>2</sub> - Second medial vein; M<sub>3</sub> - Third medial vein; PUP - Punjabi University, Patiala; R - First radial vein; R - Second radial vein; R - Third radial vein; R - Fourth radial vein; R - Fifth radial vein; Rs - Radial sector; Sc - subcostal vein; Sc+R<sub>1</sub> - Stalk of subcostal and first radial vein; UHF - University of Horticulture and Forestry.

*Plutella xylostella* Linnaeus (commonly known as Diamond back moth) (Family Plutellidae) is one of the major pests of the cruciferous crops, though its main hosts are cauliflower and cabbage. On world basis, this genus *Plutella* Schränk is reported by over forty species with *P. xylostella* Linnaeus being the only one to be recorded from South-East Asia (Robinson *et al.*, 1994). We have collected and examined 134 males and 55 females specimen from four states -- Jammu & Kashmir, Himachal Pradesh, Punjab and Uttranchal representing northwestern Siwaliks. Besides re-examination of the adult morphology, wing venation, male and female genitalia have been studied in detail to improve the diagnosis of the genus.

### MATERIALS AND METHODS

The Plutellid specimens collected with the help of light traps from four states were processed as per methodology discussed by Lindquist (1956), Tøgestad (1974), Zimmerman (1978), Mikkola (1986), and Landry and Landry (1994). The methodology involved killing (using tetrachloroethane), pinning and preservation (well fumigated wooden boxes) of adult specimens and stored in the Lepidoptera Laboratory, Department of Zoology, Punjabi University, Patiala. The standard techniques given by Zimmerman (1978) and Robinson (1976) were followed for wings preparation and genitalia dissection respectively. To write the taxonomic descriptions on various morphological characters, wing venation (Zimmerman, 1978; Park, 1995; Hodges, 1998) and external genitalia (Klots, 1970) were followed. For naming of various veins, Comstock-Needham system discussed by Miller (1970) was adopted. All illustrations were made by using a camera lucida attached to a stereoscopic light microscope (Nikon SMZ-

10, Japan). All the specimens were photographed prior to dissection.

### OBSERVATIONS

#### Family Plutellidae

Plutellidae Guenée, 1845, *Annls Soc. Ent. Fr.*, (2)3: 339 (as Plutellidi)

Type genus: *Plutella*, Schränk 1802, *Fauna Boica*, 2(2): 169.

#### Genus: *Plutella* Schränk

*Plutella* Schränk, 1802, *Fauna Boica*, 2(2): 169.

*Anadetia* Hübner, [1825], *Verz. Bekannt. Schmett.*: 405.

*Euota* Hübner, [1825], *Verz. Bekannt. Schmett.*: 408.

*Cerostoma* Latreille, [1802] sensu Stephens, 1834, *Illust. Br. Ent* (Haust.) 4: 341

*Creagria* Sodoffsky, 1837, *Bull. Soc. Imp. Nat. Moscou* 1837 (6): 94

Type species: *Phalaena xylostella* Linnaeus, 1758, *Syst. Nat.* (Edn. 10), 1: 538, by monotypy.

### Diagnosis

Head covered with somewhat erect scales; labial palpus long, upturned, second segment with tuft of long scales, third segment acute; antenna long, filiform, about <sup>3</sup>/<sub>4</sub> length of valvae; forewing characterised by a series of pale diamonds formed down its "back" by the overlapped pale markings on the dorsum when the moth is at rest; vein R<sub>4</sub> to costa, R<sub>5</sub> to termen, medial veins free, CuA<sub>1</sub> and CuA<sub>2</sub> free, CuP vestigial; hindwing with Rs to costa, M<sub>1</sub>+M<sub>2</sub> stalked, CuA<sub>1</sub> and CuA<sub>2</sub> free; male genitalia with uncus less sclerotized; socii absent; gnathos with two curved lobes; saccus present; valvae foot shaped, sclerotized except middle; aedeagus ankylosed, basally with lobes, vesica lacking cornutus; female genitalia characteristic anterior apophyses apically thin, long, broader at base; signum small or large, kite-shaped, surrounding sclerotized.

#### *Plutella xylostella* Linnaeus

*Plutella xylostella* Linnaeus, 1758, *Syst. Nat.* (Edn. 10) 1: 538. (Images 1-4<sup>w</sup>, Figs. 1-6)

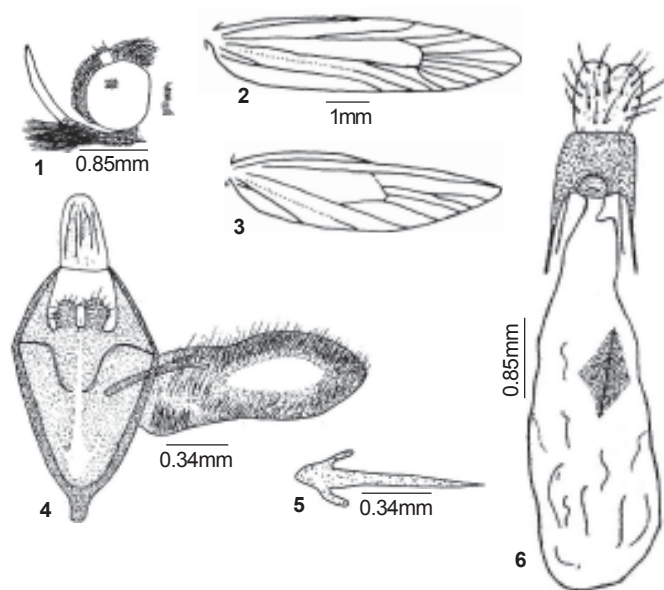
### Material examined

Four males, one female, 2.iii.1998; three males, 8.iii.1998; three males, one female, 19.iii.1998; five males, two females, 5.iv.1998; three males, two females, 7.iv.1998; three males, 11.ii.1999; two males, two females, 23.ii.1999; seven males, two females,

<sup>w</sup> see Images 1-4 in the web supplement at [www.zoosprint.org](http://www.zoosprint.org)

© Zoo Outreach Organisation; [www.zoosprint.org](http://www.zoosprint.org)

Manuscript 1322; Received 27 January 2005; Revised received 14 May 2005; Finally accepted 02 June 2005; Date of publication 21 June 2005



**Figures 1-6. *Plutella xylostella* Linnaeus**  
**1 - Labial palpus; 2 - Forewing venation; 3 - Hindwing venation; 4 - Male genitalia: ventral view; 5 - Aedeagus; 6 - Female genitalia: ventral view**

8.iii.1999; four males, two females, 15.iii.1999; three males, 23.iii.1999; seven males, three females, 29.iii.1999; seven males, two females, 6.iv.1999; two males, 1.viii.1999; two males, 17.x.1999; two males, 10.iii.2000; three males, two females, 14.iii.2000; four males, two females, 21.iii.2000; three males, three females, 24.ii.2000; three males, 27.iiiv.2000; three males, one female, 9.iv.2000; two males, one female, 2.v.2000, PUP, Dist. Patiala, Punjab, 250m; two males, 3.iv.1999; ten males, 23.iv.1999; seven males, seventeen females, 25.iv.1999; two males, 5.iv.2000; one male, 17.iv.2000; five males, 18.iv.2000; eight males, three females, 20.iv.2000, FRI Dehradun, Dist. Dehradun, Uttaranchal, 700m; two males, 18.v.1999; three males, 19.v.1999; four males, three females, 4.iv.2000, UHF Nauni, Dist. Solan, Himachal Pradesh, 1360m; three males, two females, 6.iv.2000, Dharpur, Dist. Solan, Himachal Pradesh, 1500m; two males, 12.iv.1999; two males, one female, 13.iv.1999; two males, 5.ix.1999, Renuka Lake, Dist. Sirmour, Himachal Pradesh, 740m; five males, three females, 9.xi.2001, Jammu University Campus, Dist. Jammu, Jammu & Kashmir, 350m (coll. P.C Pathania).

#### Distribution

India, Myanmar and Sri Lanka (Fletcher, 1921), world-wide including India (Vishwanath & Agarwal, 1982; Prasad *et al.*, 1983; ICAR, 1987; Velusamy *et al.*, 2000; David, 2001; Atwal & Dhaliwal, 2002), cosmopolitan (Butani & Jotwani, 1984; Butani, 1987); Europe, Asia, Africa, Western Hemisphere, Oceania (CPC, 2001).

**Biology:** Pupa (Image 1), Larva (Image 2).

**Male and female** (Images 3 & 4)

**Alar expanse:** 10-14mm. Vertex and frons covered with somewhat erect, creamish-brown scales; labial palpus moderately long,

upturned, second segment covered with creamish-fuscous tuft of long scales, third segment acute, creamish-fuscous (Fig. 1); antennae long, filiform, with white and fuscous band alternatively, about  $\frac{3}{4}$ <sup>th</sup> length of forewing; thorax creamish-brown at middle, sides fuscous in colour; forewing elongate, fuscous brown scaled, pale creamish-brown markings at base to tornus, costa slightly convex, apex acute, termen convex, tornus poorly defined, anal margin convex, termen with cilia grey; hindwing grey scaled, costa convex from base to middle then almost straight, distally convex, apex acute, termen oblique, tornus convex, anal margin convex, termen and anal margin with cilia grey in colour; prothoracic and mesothoracic legs creamish-black in colour, metathoracic leg creamish in colour.

**Wing venation:** (Figs. 2 & 3)

Forewing with Sc ending slightly below middle of costa,  $R_1$  arising slightly below middle of discal cell,  $R_1$  curved at ending,  $R_1$ ,  $R_2$ ,  $R_3$  free,  $R_2$ ,  $R_3$  parallel,  $R_3$  arising at upper angle of discal cell,  $R_4$  to costa,  $R_5$  to termen,  $M_1$ ,  $M_2$ ,  $M_3$  free, parallel,  $M_1$  arising at middle of discocellular,  $M_1$  nearer to  $R_5$  than  $M_2$ ,  $M_3$  and  $CuA_1$  arising at lower angle of cell,  $CuA_1$  and  $CuA_2$  free,  $CuA_1$  curved at middle,  $CuA_2$  arising at  $\frac{4}{5}$ <sup>th</sup> of discal cell,  $CuP$  vestigial, visible at  $\frac{1}{4}$ <sup>th</sup> distally,  $1A+2A$  forked at base; hindwing with  $Sc+R_1$  ending near middle of costa,  $Rs$  to costa, straight, arising at upper angle of cell,  $M_1+M_2$  stalked, arising at middle of discocellular,  $M_3$  free, near to  $M_2$  than  $CuA_1$ ,  $CuA_1$ ,  $CuA_2$  free,  $CuA_1$  arising at lower angle of cell,  $CuA_2$  arising at  $\frac{2}{3}$ <sup>rd</sup> of discal cell,  $CuP$  vestigial, visible at ending,  $1A+2A$  small forked at base.

**Male genitalia** (Figs. 4 & 5)

Uncus not sclerotized; socii absent; gnathos with two curved lobes, small, sparsely setose, joint with tegumen; tegumen small, somewhat arched; juxta weak and thin; vinculum long and broad, Ushaped; saccus small, rounded apex; valvae symmetrical, foot-shaped, well sclerotized except at middle, inner margin densely setose, costal margin slightly convex, with a long arm at base, directed posteriorly, sacculus margin concave apically, convex basally with bunch of spine near base, cucullus margin strongly convex dorsally, apex rounded, convex ventrally; aedeagus small, ankylosed, broader at base then gradually narrowed, pointed at apex, two small lobes near base; vesica without cornutus.

**Female genitalia** (Fig. 6)

Papillae anales large and broad, sparsely setose; anterior apophyses small, broader at base, apically thin, posterior apophyses long and thin, longer than the length of anterior apophyses; ostium bursae small opening, side sclerotized, centrally placed; ductus bursae very small, somewhat broad; corpus bursae long and broad, broader distally then gradually narrowed upto base; signum small, kite-shaped, moderately sclerotized.

**Larval host plant:** Cabbage, cauliflower, raddish, mustard and cruciferous plants (Fletcher, 1921).

**REMARKS**

Bradley (1966) designated a lectotype of the species, *Phalaena xylostella* Linnaeus, 1758 and in view of this *P. xylostella* was placed on the official list of specific names in zoology (No. 2506). While giving a key to the Philippine genera of the family Plutellidae, Diakonoff (1967) has stressed that the genus, *Plutella* Schrank can be distinguished on the basis of the veins  $M_1$  and  $R_3$  (Figs. 2, 3) of the forewing, which are always present and separate. It can be characterised by the series of pale diamonds formed down its back by the overlapped pale markings on the forewing dorsum, when the moth is at rest. The second segment of the labial palpi is conspicuous due to the presence of a porrect tuft of very long scales on it.

**REFERENCES**

- Atwal, A.S. and G.S. Dhaliwal (2002).** *Agricultural pests of South Asia and their management*. Kalayani publishers, New Delhi, 498pp.
- Bradley, J.D. (1966).** A comparative study of the coconut flat moth (*Agonoxena argaula* Meyrick) and its allies, including a new species (Lepidoptera, Agonoxenidae). *Bulletin of Entomology Research* 56: 453-472 + 1-38 figs.
- Butani, D.K. (1987).** Key pests of vegetables and their management, pp. 433-454. In: Mathur, Y.K., A.K. Bhattacharya, N.D. Pandey, K.D. Upadhyaya and J.P. Srivastava (Eds), *Recent advances in entomology*. Gopal Prakashan Parade, Kanpur.
- Butani, D.K. and M.G. Jotwani (1984).** *Insects in vegetables*. Periodical Expert Book Agency. New Delhi, 356pp.
- CPC (2001).** *Crop Protection Compendium*. CAB International, 2001 Edition, ISSN: 1365-9065.
- David, B. V. (2001).** *Elements of economic entomology*. Popular Book Depot. Chennai, 562pp+28pls.
- Diakonoff, A. (1967).** Microlepidoptera of the Philippine Islands. *Bulletin of U.S. natn. Museum* 257: 1-484.
- Fletcher, T.B. (1921).** Life history of Indian Insects, Microlepidoptera. *Memoirs of Department of Agriculture India* 6: 1-217.
- Hodges, R.W. (1998).** *Gelechioidea*, pp. 130-158. In: Kristenstensen, N.P. (ed.), *Lepidoptera, Moths and Butterflies*. Handbook of Zoology/Handbuch der Zoologie 1 (1999). Berlin and New York.
- ICAR, (1987).** *Hand Book of Agriculture*. Indian Council of Agricultural Research, New Delhi, 1393pp.
- Klots, A.B. (1970).** Lepidoptera, pp. 97-111+121-132figs. In: Tuxen, S.L.(ed.) *Taxonomists Glossary of Genitalia in Insects*. Copenhagen, 284pp.
- Landry, J.F. and B. Landry (1994).** A technique for setting and mounting microlepidoptera. *Journal of Lepidoptera Society* 48(3): 205-227.
- Lindquist. O.H. (1956).** A technique for pinning and spreading small microlepidoptera. *Canadian Entomology* 88(1): 24-25.
- Mikkola, K. (1986).** Tower spreading, a handy method for provisional field preparation for microlepidoptera. *Not. Entomology* 66: 101-102.
- Miller, L.D. (1970).** Nomenclature of wing veins and cells. *Journal of Research in Lepidoptera* 8(2): 37-48.
- Park, K.T. (1995).** Gelechioidea of Taiwan I. Review of *Anarsia* with descriptions of four new species (Lepidoptera: Gelechioidea). *Tropical Lepidoptera* 6(1): 55-66.
- Prasad, V. G., K. Krishnaiah and Jagan Mohan (1983).** Insect pests of vegetables, pp. 235-260. In: Srivastava, P.D., M.G. Jotwani, R.A. Agarwal, S.R. Wadhi, R.K. Bhanotar and R.K. Bhatnagar (Eds.), *Agricultural Entomology Vol. II*. All India Scientific Writers Society, New Delhi,
- Robinson, G.S. (1976).** The preparation of slides of Lepidoptera genitalia with special reference to microlepidoptera. *Entomologist Gazette* 27(2): 127-132.
- Robinson, G.S., K.R. Tuck and M. Shaffer (1994).** *A field guide to the smaller moths of South-East Asia*. Malaysian Nature Society, Malaysia, 1308pp+132pls.
- Tagestad, A.D. (1974).** A technique for mounting microlepidoptera. *Journal of Kansas Entomology Society* 47: 26-30.
- Velusamy, R., P. Sivasubramaniam, S. Suresh and R. Rajendran (2000).** *Integrated pest management in horticultural crops*. Department of Entomology, AD AC and RI, Trichy, 158pp.
- Vishwanath, J. and R.A. Agarwal (1982).** *Insect pests of crops and their control*. Bharati Publications, New Delhi, 139pp.
- Zimmerman, E.C. (1978).** *Microlepidoptera. Insects of Hawaii, 9*. University Press of Hawaii, Honolulu, 1903pp+xvii.

**ACKNOWLEDGEMENTS**

We are grateful to the Ministry of Environment and Forests (GOI), New Delhi for funding the project on taxonomy of Microlepidoptera. The authors are also thankful to Dr. K.T. Park (Korea) and Dr. G.S. Robinson, (N.H.M.), London for sending much needed literature.



## The 2nd Conference of the International Congress of Zookeepers

Gold Coast, Queensland, Australia  
7th – 11th May 2006

### Second Call for Papers/Posters and Workshops

After the success of the 1<sup>st</sup> conference in 2003, a second conference will be held in Australia in 2006 with more than 300 zookeepers from around the world expected to attend. The conference will cover all areas of Zookeeping.

You are invited to submit an abstract of a paper or presentation you would like to give. More information and guidelines can be found at [www.iczoo.org](http://www.iczoo.org)

Abstracts for oral and poster presentations should be written in English and no more than 600 words long. If you want to run a workshop focused on developing zoo keeper skills please send a short description.

These should be sent to: Paul Howse, ICZ Programme Committee at [p.howse@chesterzoo.org](mailto:p.howse@chesterzoo.org)

Preferably in RTF or MS word format marked 'ICZ Programmme'.

**Deadline for submissions is 30 September 2005**